

HIGH PRODUCTIVITY PROTEIN A MEMBRANE DEVICES COMPLEMENT DISPOSABLE UPSTREAM TECHNOLOGY FOR A FULLY SINGLE-USE PROCESS

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Key Trends Impacting Bioprocessing Today

SINGLE-USE SYSTEMS

- Smaller footprint, less capital
- Sustainability: less water, chemicals and energy
- Scalable & flexible
- Less downtime & faster turnover
- Lower risk of contamination, bioburden, and operator error

INTENSIFIED PROCESSING

- Smaller footprint producing higher productivity
- Consistent scalability & facility fit to speed tech transfer

SUPPLY CHAIN

- Dual source options
- Short lead times

NEW, COMPLEX MODALITIES

- Multispecifics, ADCs, Fc fusions, etc.





The Downstream Bottleneck: Resin Chromatography

- ⊗ Protein A resin columns are **oversized/underutilized** to gain flow and speed processing times to keep pace with upstream productivity
- ⊗ **Need to optimize chromatography** media performance to enable high flow rate and high binding capacities
- ⊗ **Facility fit challenges** for multiple drugs, scales and potencies
- ⊗ **Downtime** for packing, qualifying, storing, validating resin columns
- ⊗ **Bioburden** that can impact your bottom line and your project timeline

Proprietary Membrane, Immobilized Protein A

IMPROVES PRODUCTIVITY

- Consistent scaling, speed, capacity, elution width and pressure drop

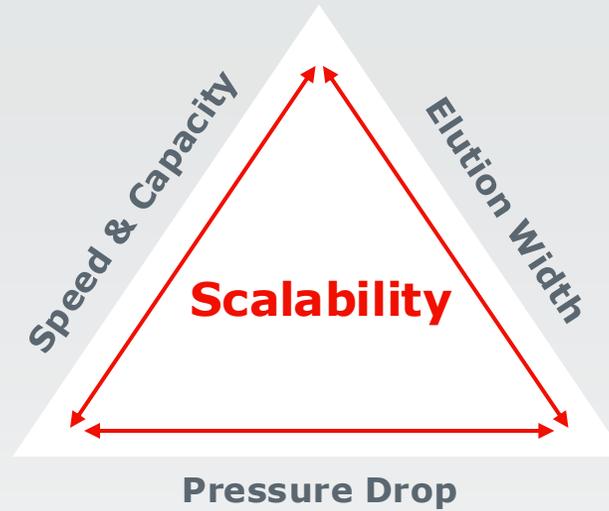
RAPID CYCLE AND FLEXIBLE PROTOCOLS

COMMERCIALLY AVAILABLE PREPACKAGED COLUMN SIZES

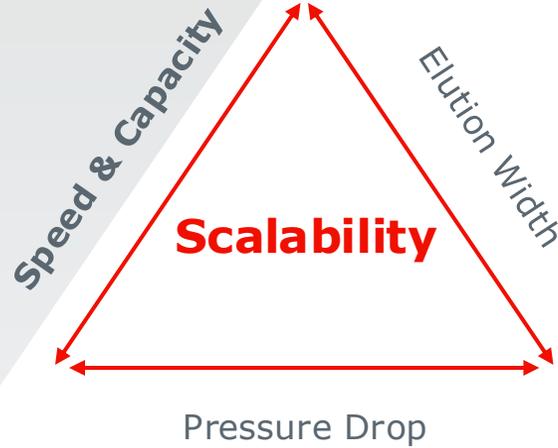
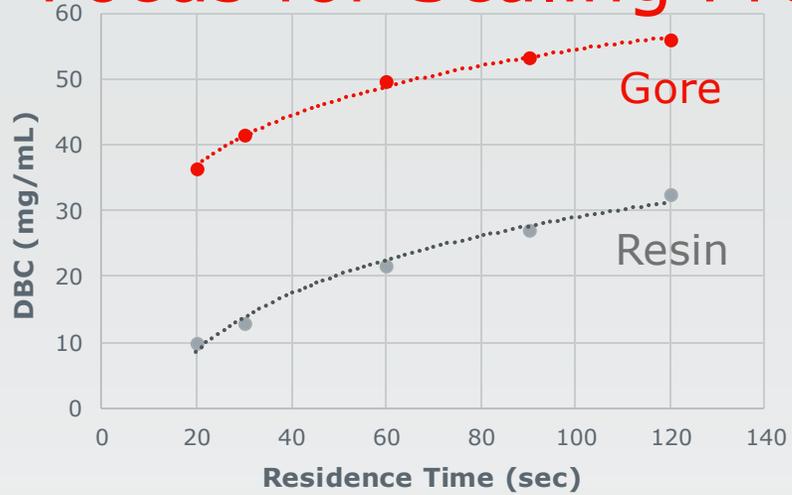
- Lab scale: 1.0 mL and 3.5 mL
- GMP supported: 9.0 mL, 58 mL, 116 mL, 250 mL, 500 mL, and 1L (GMP)



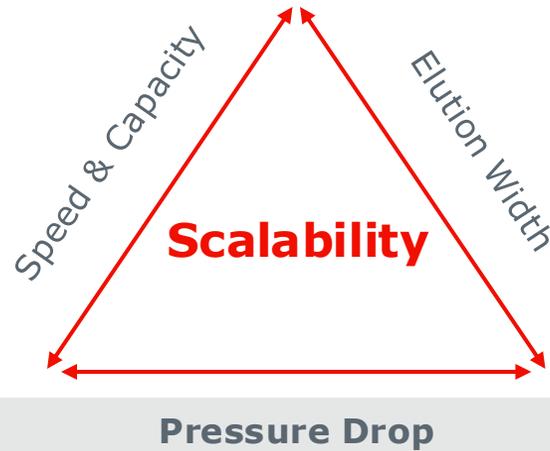
Focus for Scaling Proprietary Membrane



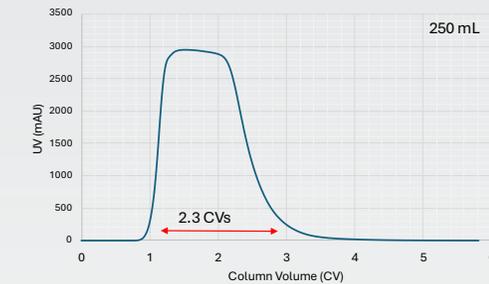
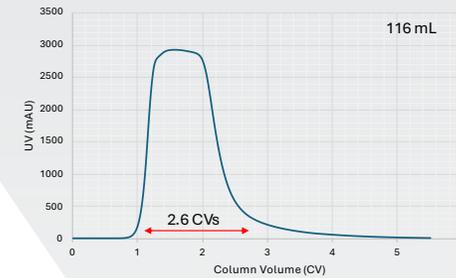
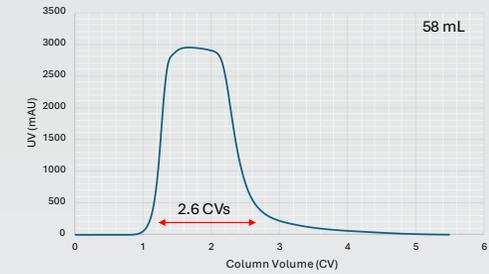
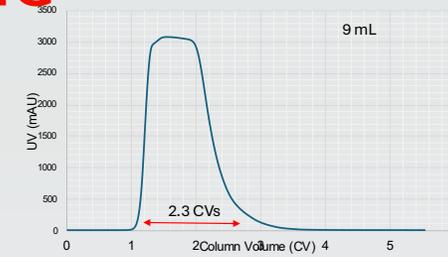
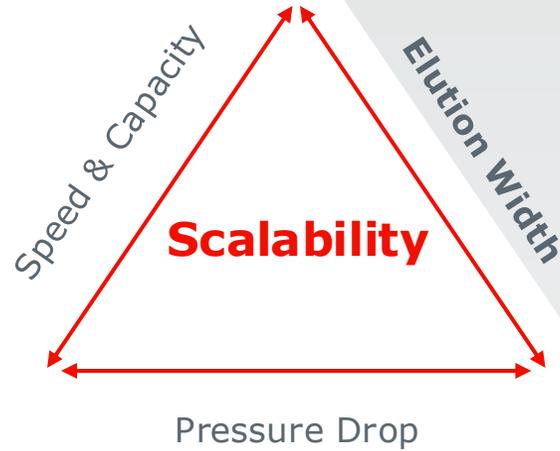
Focus for Scaling Proprietary Membrane



Focus for Scaling Proprietary Membrane

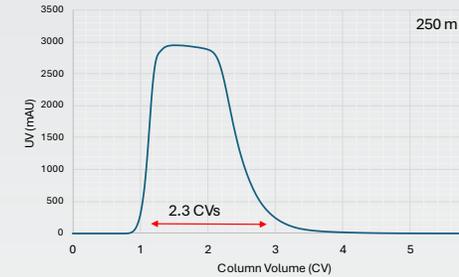
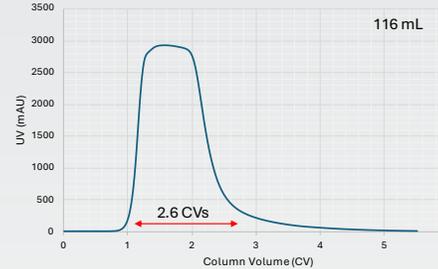
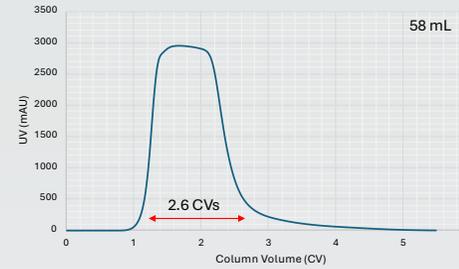
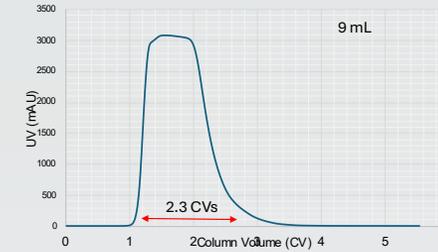
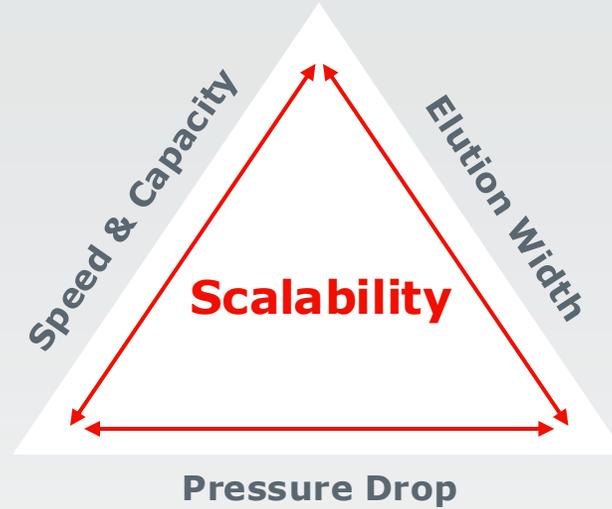
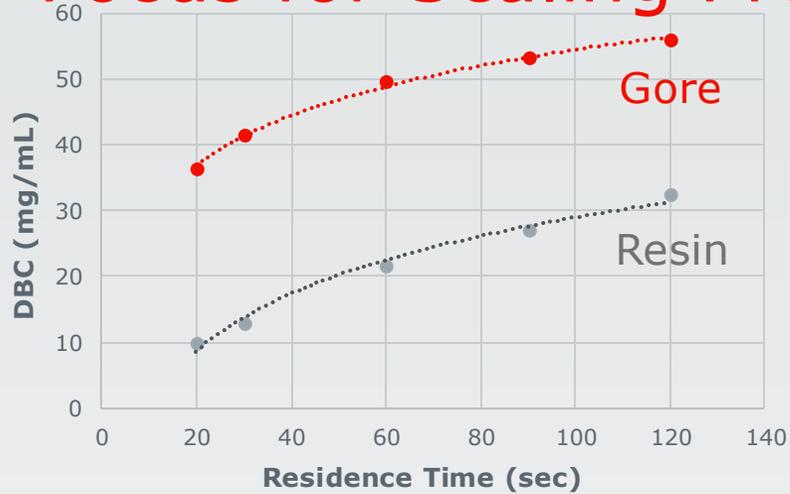


Focus for Scaling Proprietary Membrane



Elution Width on average ~ 2.5 MV

Focus for Scaling Proprietary Membrane

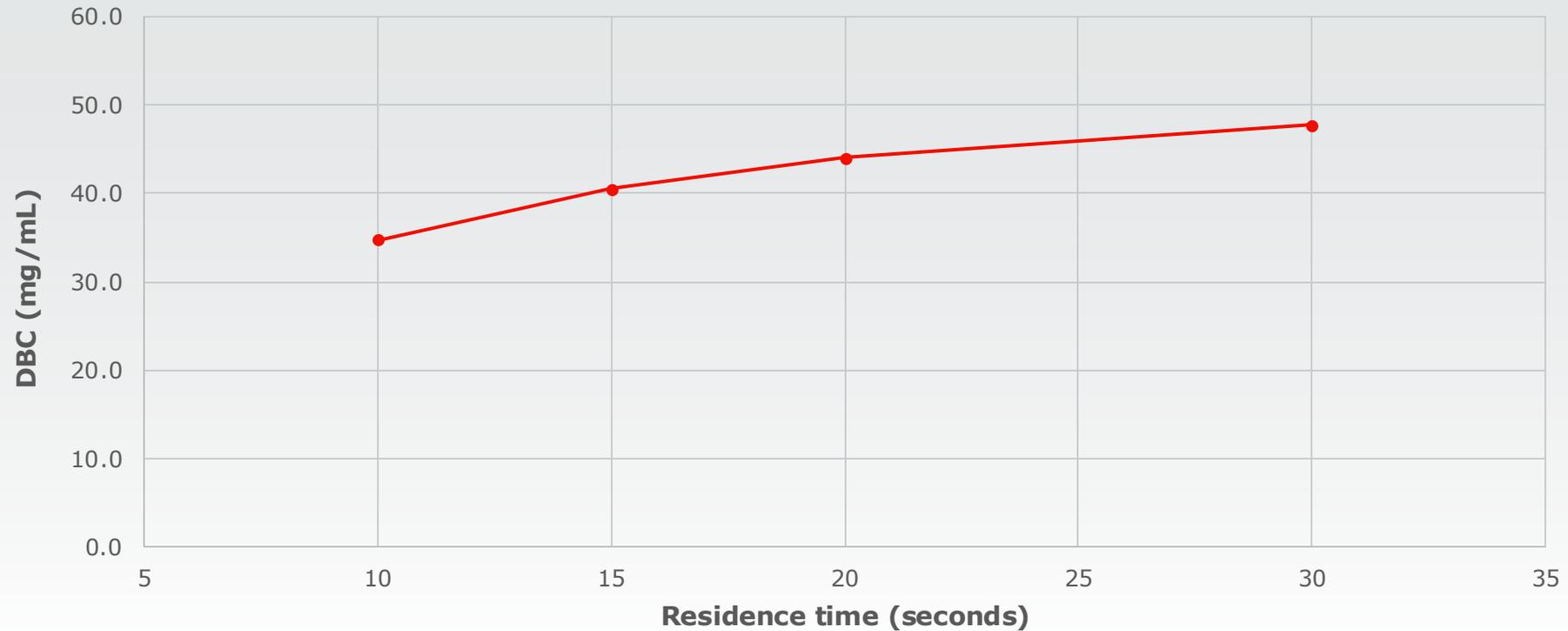


Elution Width on average ~ 2.5 MV

Focus on Productivity Optimization

Faster than 20srt

Gore® Protein Capture Device Dynamic Binding Capacity
(hIgG at 10% Breakthrough)



Improved Productivity with Flexible Protocols

GORE® Protein Capture Devices with Protein A — 58 mL: 4g/L CHO cell harvest

Initial Study:
88 g L⁻¹ h⁻¹

- Loading: 30srt
- Operational: 10srt
- Conservative transitions
- 3 minute CIP per cycle
- 16.8 minutes/per cycle

Reduce Transitions:
143 g L⁻¹ h⁻¹

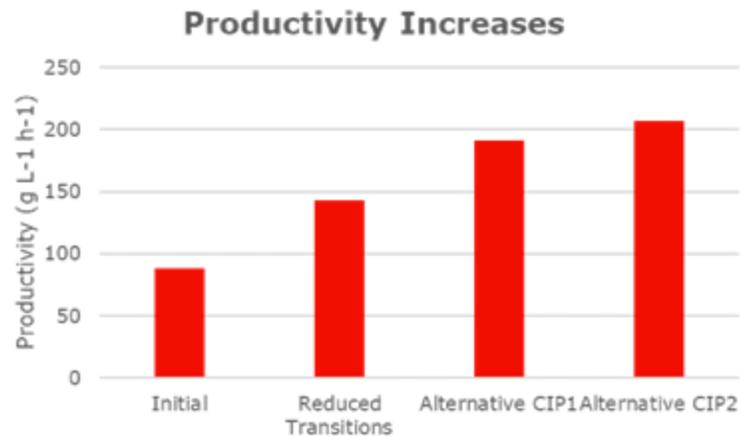
- Loading: 30srt
- Operational: 10srt
- Reduced transitions
- 3 minute CIP per cycle
- 12.6 minutes/per cycle

Alternative CIP1:
191 g L⁻¹ h⁻¹

- Loading: 15srt
- Operational: 7srt
- Reduced transitions
- 2 minute CIP per cycle
- 9.5 minutes/per cycle

Alternative CIP2:
207 g L⁻¹ h⁻¹

- Loading: 15srt
- Operational: 7srt
- Reduced transitions
- "Pulse CIP" per cycle
- 6.9 minutes/per cycle

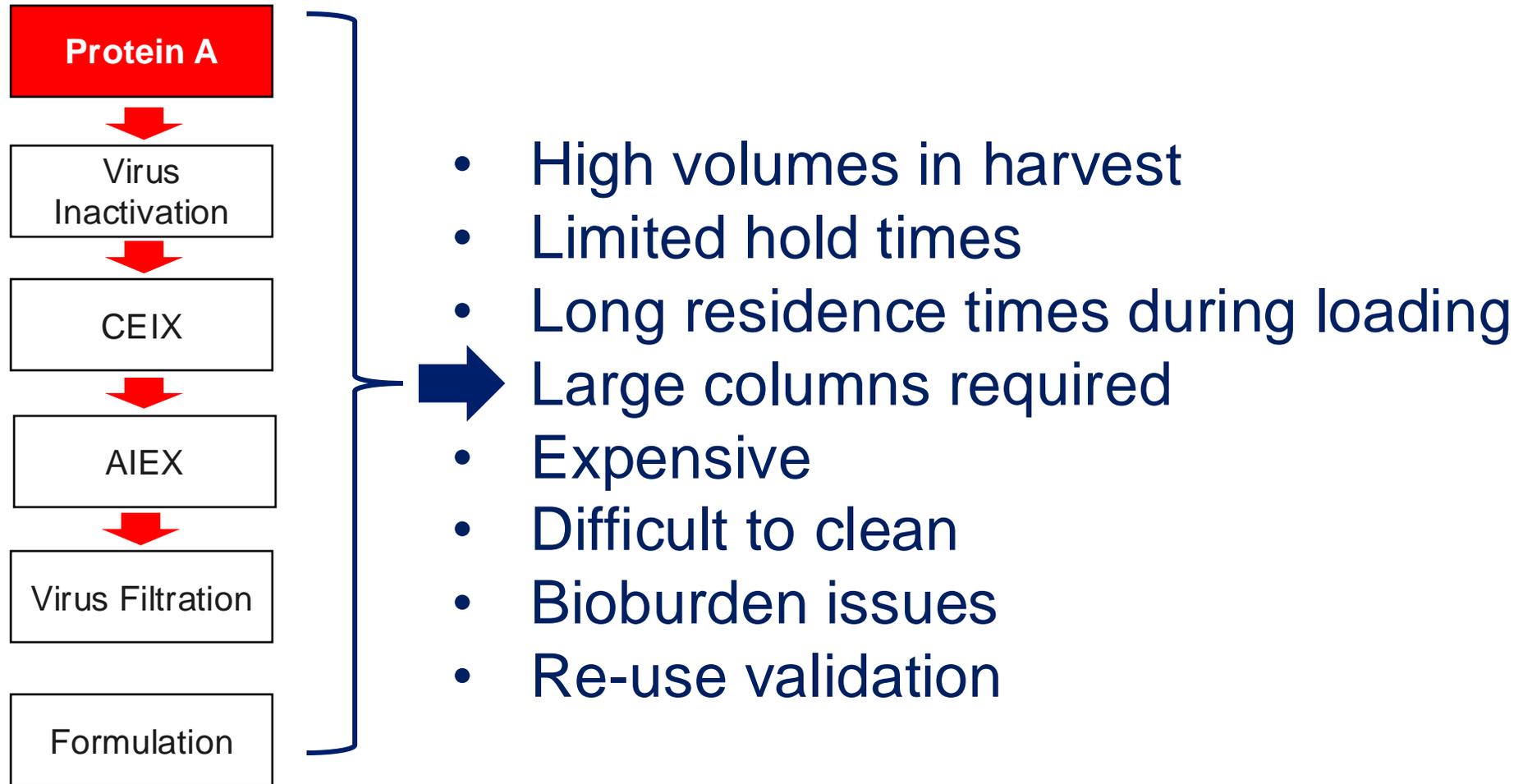


- CMO and pharma evaluations show no difference in product quality attributes relative to productivity
 - HCP clearance
 - Yield
 - Protein A leaching
- Similar CQAs to resin
- Validated through 100 cycles and demonstrated 200 cycles
- Flexible CIP and durability enable ability to clear 2000-L bioreactor in fully disposable process



Results from Partner Research: Single-Use Upstream + Protein A Purification Devices

Protein A Resin Limitations in Standard mAb Platforms



Gore Protein A Membrane | 1 L Manufacturing Scale Device



2 x 1 L Protein A Devices



The Research Execution

- Scale-up run using 2 x 1 L Gore Protein A devices
- Small footprint required (approx., 10 x 10 inch for each column).
- Standard single use chromatography skid used

Protein A Membrane | Short Residence Time and Rapid Mass Transfer

Harvest titer : 1.717 g/L
 Membrane volume : 2 L
 Load volume: 23.5 L (40 g/cycle)

We performed two method to reduce sanitization volume between cycles

Cycle 1-10

| Step | Volume (MV) | Residence Time (min) | Total Time (min) |
|----------------------------|-------------|----------------------|------------------|
| Equilibration | 3.0 | 0.2 | 0.6 |
| Load | 11.6 | 0.4 | 4.66 |
| Wash 1(EQ wash) | 1.43 | 0.4 | 0.57 |
| Wash 2(Salt wash) | 3 | 0.2 | 0.60 |
| Pre-elution wash (EQ wash) | 3 | 0.2 | 0.60 |
| Elution | 3.75 | 0.2 | 0.75 |
| Sanitization (0.1N NaOH) | 5 | 0.4 | 2.00 |
| Re-Equilibration | 3 | 0.2 | 0.60 |
| Total | 33.8 | - | 10.38 |

Cycle 11-19

| Step | Volume (MV) | Residence Time (min) | Total Time (min) |
|----------------------------|-------------|----------------------|------------------|
| Equilibration | 3.0 | 0.2 | 0.6 |
| Load | 11.6 | 0.4 | 4.66 |
| Wash 1(EQ wash) | 1.43 | 0.4 | 0.57 |
| Wash 2(Salt wash) | 3 | 0.2 | 0.60 |
| Pre-elution wash (EQ wash) | 3 | 0.2 | 0.60 |
| Elution | 3.75 | 0.2 | 0.75 |
| Sanitization (0.1N NaOH) | 3 | 0.4 | 1.20 |
| Re-Equilibration | 3 | 0.2 | 0.60 |
| Total | 31.8 | - | 9.58 |

Protein A Membrane | A280 Chromatograms & High Reproducibility

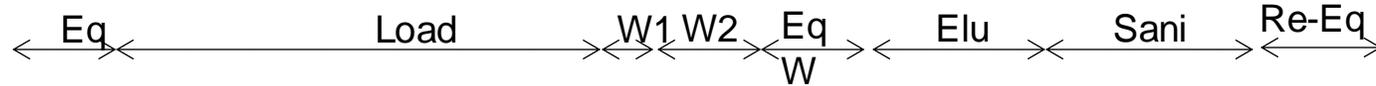
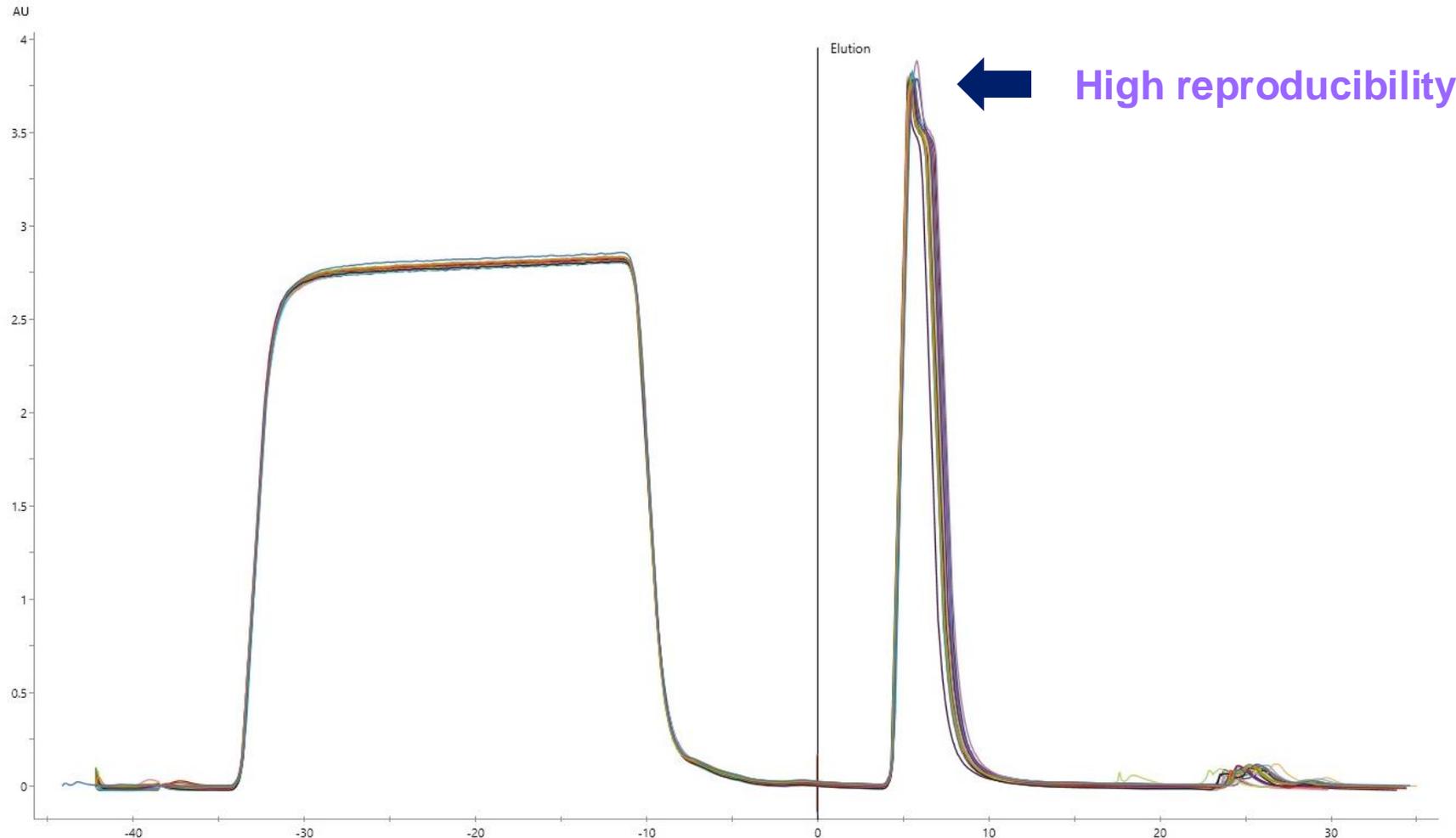
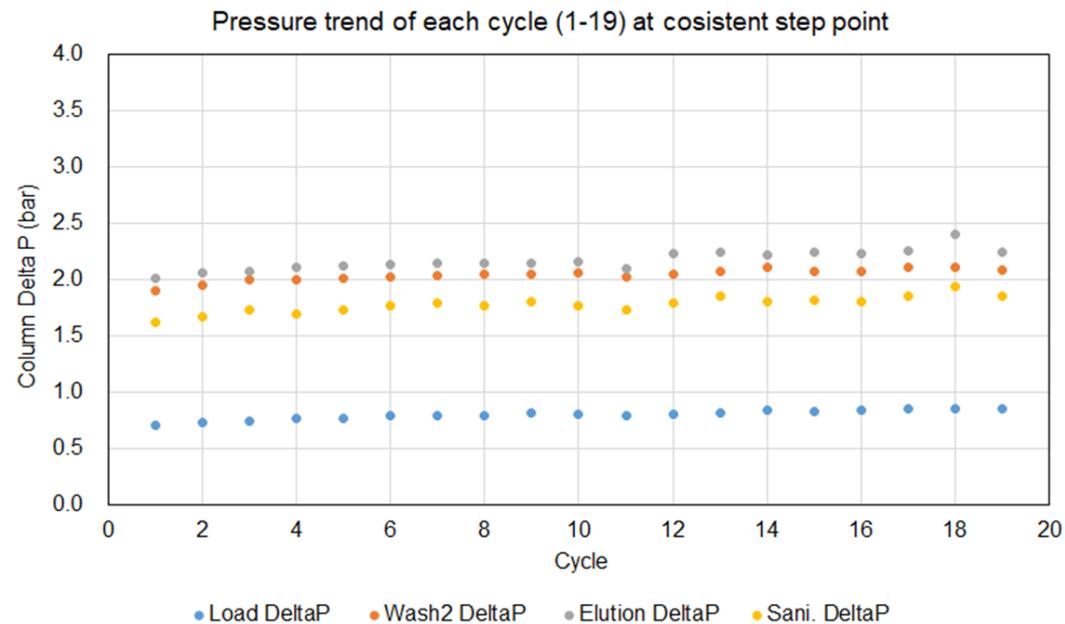
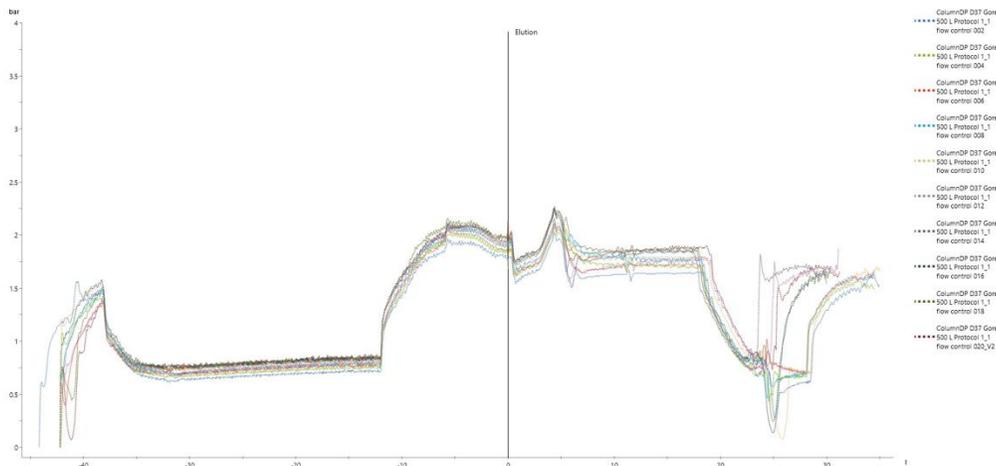


Diagram: Overlay of A280 Chromatograms for each cycles 1-19



Protein A Membrane at 500L | Pressure Trends



Maximum operating pressure : 4 bar

Over 19 cycles:

- No clogging of the device occurred and there was no sudden increase in pressure
- There was no increase in pressure when Sani volumes were reduced (after Cycle 11)

Lab Scale Results: Protein A Resin vs. Gore Device Demonstrate Good Performance and Quality

| | 5 mL Protein A Resin column | 3.5 mL Gore Device with Protein A | 2 L Gore Device with Protein A |
|--|-----------------------------|---|--|
| Productivity (g/L/h) | 13.8 | 132.8  | 132.6 |
| Average elution volume [100-100 mAU cutoff] | 2.72 | 1.84 | 2.68 |
| Elution HCP (LRV) | 2.03 | 2.12 | 2.22  |
| Elution Protein A (ppm) | 4.28 | 5.25 | 2.49 |
| Average SEC Product Quality (% main/%HMW) | 89.9 / 10.1 | 93.3 / 6.7 | 93.9 / 6.1 |

Notes:

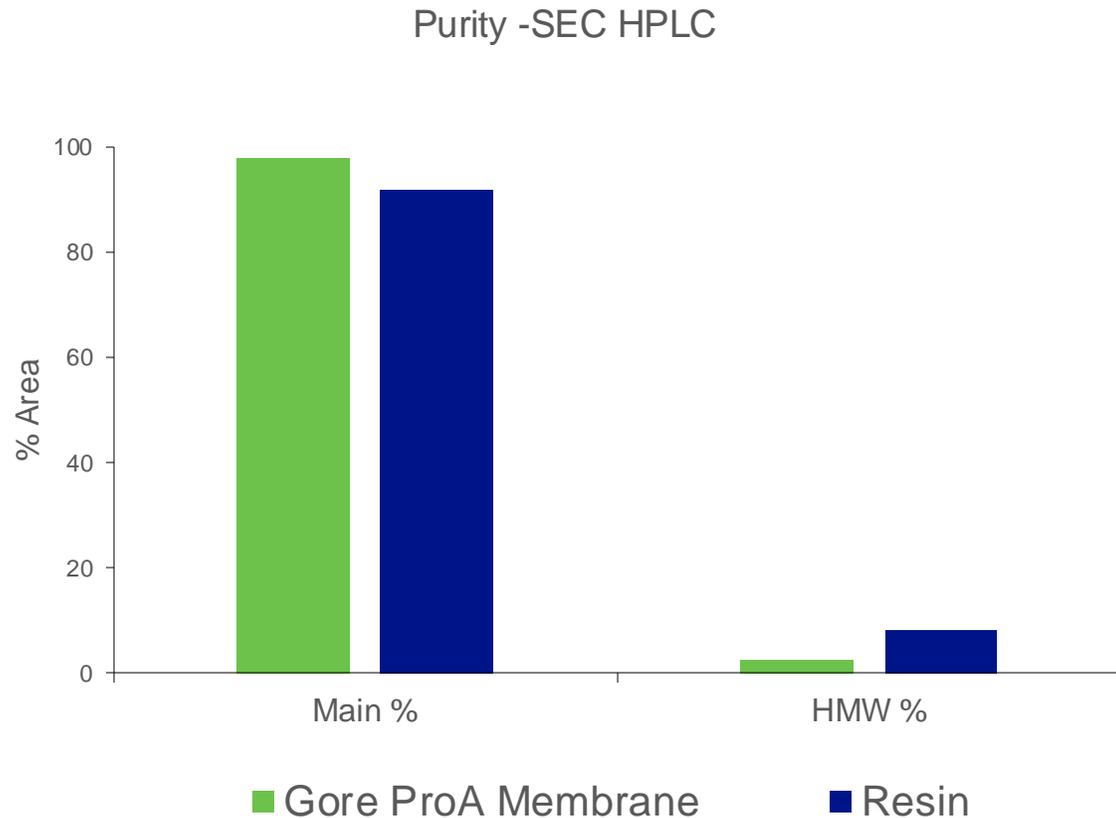
Cell culture harvest purified with laboratory-scale columns to provide benchmark performance targets & scaling reference for the Gore Protein A membrane device.

Productivity calculations indicate a consistent **10X increase in productivity** for the Gore Protein A membrane devices compared to the resin column.

Chromatographic performance and process and product impurity data were comparable between the two Gore Protein A membrane devices and the Protein A resin control.

Consistent scaling in membrane makes the process **easy to transfer across sizes** based on residence time

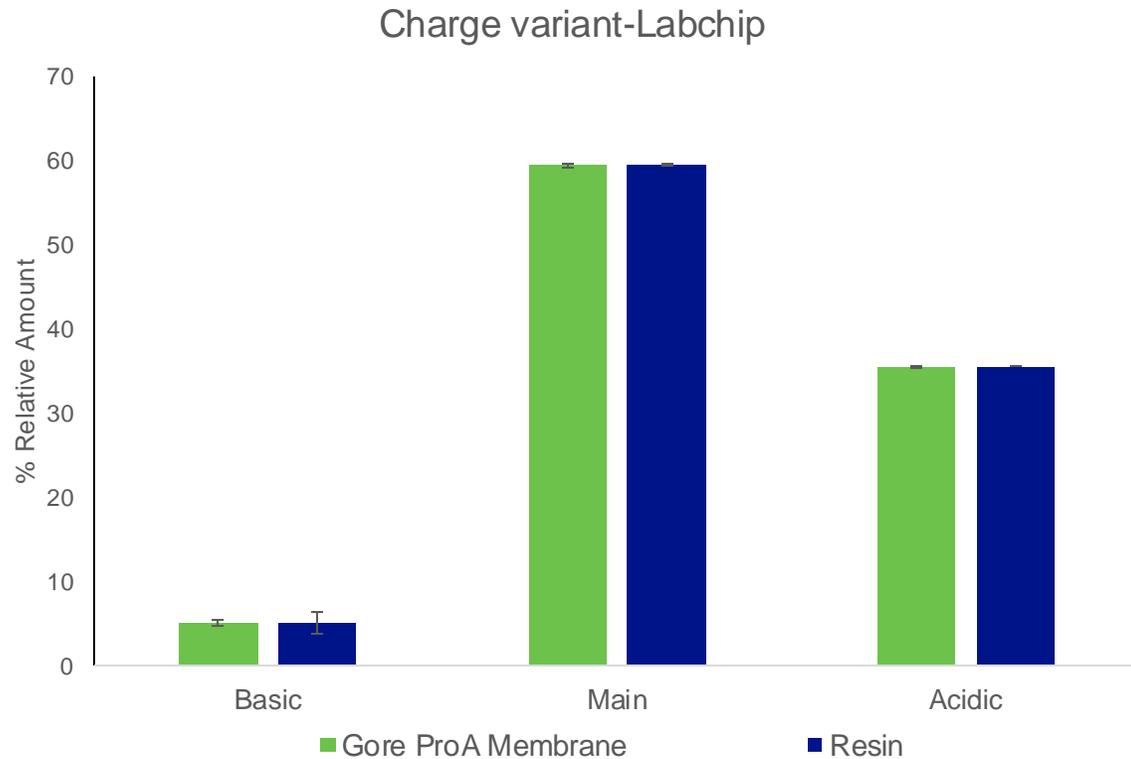
Demonstrating Similarity Membrane to Resin – Lab Scale



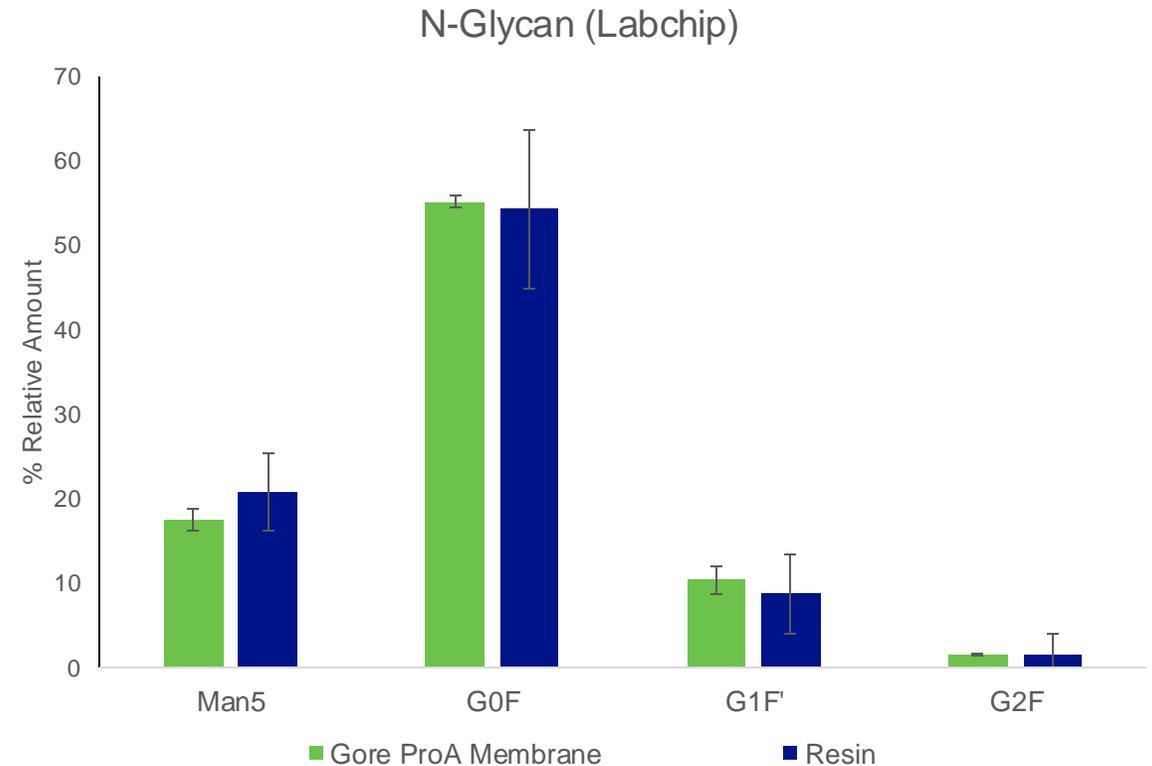
- Eluent Average SEC Product Quality distributions for laboratory scale Protein A resin column and Gore Protein A membrane device
- LMW species, indicative of protein fragmentation, were typically < 0.04 area percent in all cases

Demonstrating Similarity Membrane to Resin – Lab Scale

Eluent Charge Variant distributions for Laboratory scale Protein A resin column and Gore Protein A membrane device*.



Eluent n-glycan distributions for Laboratory scale Protein A resin column and Gore Protein A membrane device*.



*Error bars represent relative standard deviation on replicate preparations

Single-Use Downstream Performance Characterization Summary

| Step | Step Yield (%) | Step Productivity | HCP | | Residual Protein A [ppm] | Host Cell DNA | SEC-HPLC (%) | |
|---------------------------------------|----------------|---|---------|----------|--------------------------|---------------|--------------|------|
| | | | ppm | Step LRV | | Step LRV | Main | HMW |
| Harvest (Filtration) | - | - | 160,187 | - | - | - | - | - |
| Protein A Affinity (Gore Protein A) | 103.8 | 132.6 g L ⁻¹ h ⁻¹ | 830 | 2.29 | 2.49 | 3.37 | 93.9 | 6.11 |
| Viral Inactivation (Single-use Mixer) | - | - | 915 | - | 2.36 | - | 92.6 | 7.37 |
| Anion Exchange (Sartobind Q) | 88.9 | 189.2 g L ⁻¹ h ⁻¹ | 72 | 1.10 | 2.20 | 1.70 | 92.1 | 7.89 |
| TFF: UF/DF (Pellicon Capsule) | 104.4 | 72.0 g m ⁻² h ⁻¹ | 99 | - | 2.42 | - | 93.9 | 6.05 |

Table summary:

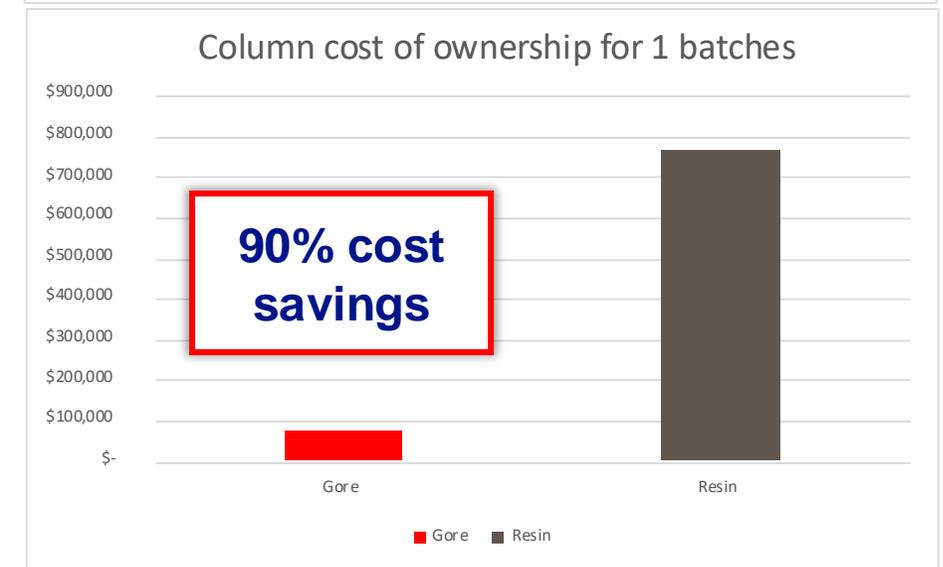
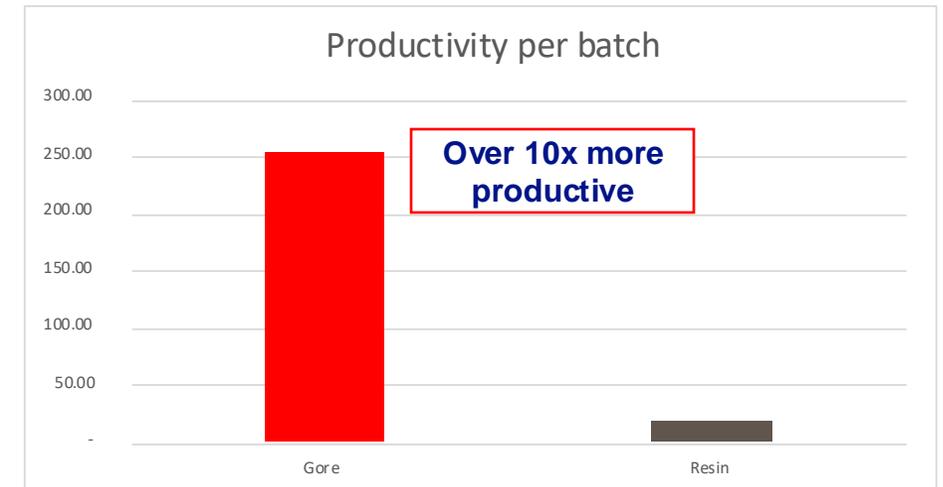
1. Step yields, step productivities, and pooled HCP, Protein A and SEC characterization data for the affinity capture, VI, AEX and TFF steps.
2. Comparison of step yield, productivity, and process/product quality data for the downstream unit operations. Purity data were analyzed from the final pool at each operation.

Potential for 90% Cost Savings, More Productivity and Faster Preparation

Assumptions made in calculations:

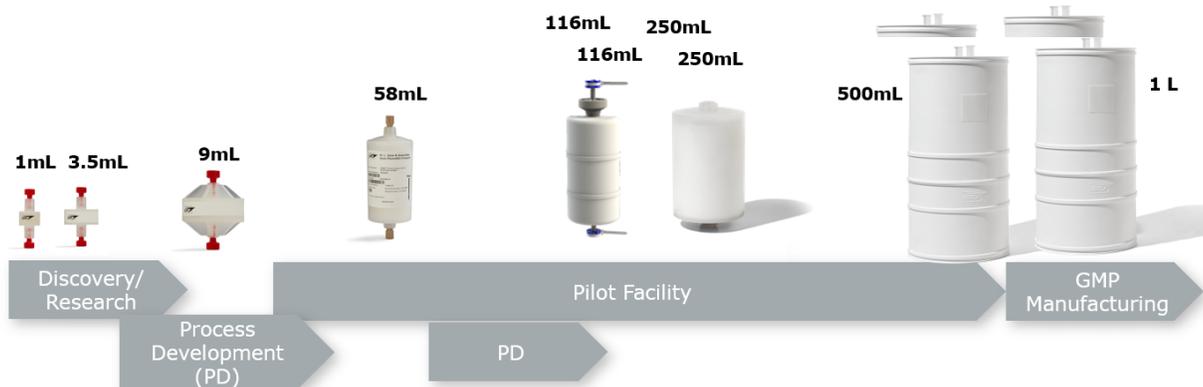
- 2,000 L Bioreactor | 5 g/L | 1 batch example | Resin: \$18,000/L

| | Resin Column | Gore Protein A Membrane Column | |
|----------------------------|--|--|-----------------------------|
| Column size | 32 L packed bed | 2 x 1 L | More efficient facility fit |
| Cost for 1 batch | \$767,900* | \$76,400 | |
| Residence/Step time | 4 minute residence time | Rapid cycle (30 seconds load, 7 seconds operational, 30 seconds CIP) | |
| Cycles per batch | 8 cycles/batch | 157 cycles/batch | |
| Processing time | 17.5 hours | 19.6 hours | Similar unit op. time |
| Preparation time | 32 hours* <small>*Assumes full Resin packing of empty column (23 hours) + setup (6 hours)</small> | 2 hours | 94% faster preparation |
| Storage cost | 10% Resin cost | None | |



Scalability of GORE Protein Capture Device

Lab Scale to GMP (2000-L focus)



| 1mL | 3.5mL | 9mL | 58mL | 116mL | 250mL | 500mL | 1L |
|----------|----------|----------------|--------------|-------------------------|-------------------------|-------------------------|-------------------------|
| R&D | R&D | PD | Pre-clinical | Pre-clinical & clinical | Pre-clinical & clinical | Pre-clinical & clinical | Pre-clinical & clinical |
| Early PD | Early PD | Late Phase R&D | Tox | Tox | Tox | Tox | Tox |
| | | GMP | PD | PD | PD | PD | PD |
| | | | GMP | GMP | GMP | GMP | GMP |

| Bioreactor Batch Size (L) | Titer (g/L) | Gore Protein A Membrane Device Configuration | Cycles ² Needed to Clear Bioreactor | Total Time per Cycle ³ (min) | CIP Time per Cycle (min) | Total Bioreactor Clearance Time (h) | Productivity (g/L/h) ⁴ |
|---------------------------|-------------|--|--|---|--------------------------|-------------------------------------|-----------------------------------|
| 2000 | 1.5 | 1 L | 94 | 14.8 | 0.5 | 23.2 | 130 |
| | | | | 16.3 | 2 | 25.5 | 118 |
| | | | | 17.3 | 3 | 27.1 | 111 |
| 2000 | 3 | 2 x 1 L parallel manifold | 94 | 9.5 | 0.5 | 14.8 | 203 |
| | | | | 11.0 | 2 | 17.2 | 175 |
| | | | | 12.0 | 3 | 18.7 | 160 |
| 2000 | 6.4 | 4 x 1 L parallel manifold | 100 | 6.7 | 0.5 | 11.1 | 289 |
| | | | | 8.2 | 2 | 13.6 | 236 |
| | | | | 9.2 | 3 | 15.3 | 210 |
| 2000 | 10 | 4 x 1 L parallel manifold | 157 | 5.8 | 0.5 | 15.0 | 334 |
| | | | | 7.3 | 2 | 18.9 | 265 |
| | | | | 8.3 | 3 | 21.5 | 233 |
| | | | | 10.3 | 3 | 26.7 | 187 |

¹Assumes DBC_{10%} of 40 g/L at 30 SRT, loading to 80% of DBC_{10%} at 30 SRT, non-loading steps at 7 SRT.

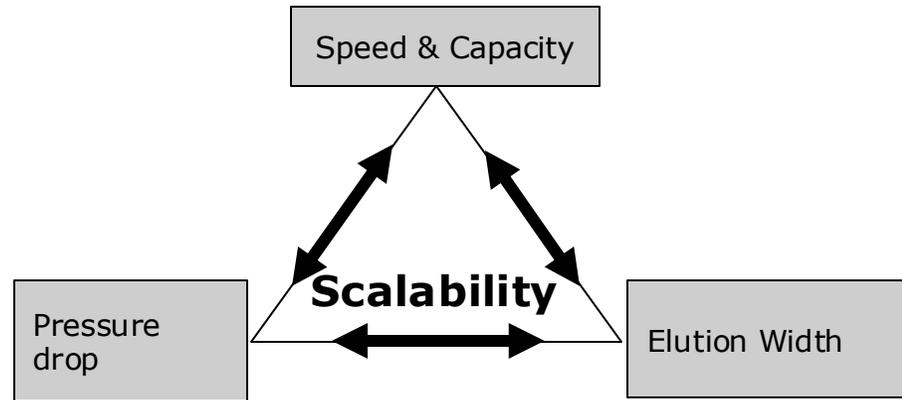
²GORE Protein Capture Devices with Protein A are validated to 100 cycles and demonstrated to 200 cycles.

³Range of total time per cycle is inherently higher for lower titers due to increased time needed for loading low titer harvest to 80% of DBC_{10%}

⁴The Gore Productivity Calculator (Gore document PB11711) can be used to model a wide range of additional scenarios

Conclusions & Looking Forward

- Consistent device performance from **1mL** to **1L**
 - ✓ Elution widths
 - ✓ Cycling purity & yield
 - ✓ Elution widths
 - ✓ Low pressure drop



Increase productivity through High binding capacity, short residence time, and low pressure drop.

THANK YOU

Visit Booth 310

W. L. Gore & Associates, Inc.

For more information:

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- Thanks to AGC Biologics Bothell and Longmont Teams

Together, improving life

